# ROLE OF FUNGI IN BIODEGRADATION OF XENOBIOTIC COMPOUNDS IN THE ENVIRONMENT

#### SUBHASH CHANDRA

P.G. Department of Botany, Sri Murli Manohar Town P G College Ballia (U. P.), India Email: subhashcbsi1@gmail.com

## **ABSTRACT**

The term biodegradation means transformation of a chemical compound from highly complicated form (organic) to simple (inorganic) form through biological process. If we say a compound is biodegradable, it means that it can be converted into various inorganic forms or can be mineralized i. e., possible to convert into carbon dioxide and water. Several species of fungi like Trichosporon cutaneum, Candida lipolytica, Candida tropicalis, Candida parapsilosis, Candida ernobii Candida boidinii Rhodotorula rubra, Aureobasidion (Trichosporon) pullulans. Rhodotorula aurantiaca, Saccharomyces cerevisiae Rhodotorula pilimanae, Yarrowiali polytica, Hansenula polymorpha Cladosporium, Aspergillus ustus, Aspergillus sydowii and Aspergillus destruens, whereas fungi belonging to Cunninghamella, Penicillium, Fusarium Cephalosporium Alternaria alternate, Mucor racemosus, Phoma glomerata, Trichoderma longibrachiatum and Trichoderma hamatum play a major role in the process of biodegradation of xenobiotic compounds in the environment.

**Key-words:** Fungi, xenobiotic compounds, biodegradation, environment

## INTRODUCTION

Biodegradation is the process by which organic substances are broken down into simple compounds by living microorganisms (Marinescu, *et al.*, 2009). When biodegradation is complete, the process is called mineralization. However, in most cases the term biodegradation is generally used to describe almost any biologically mediated change in a substrate (Bennet, *et al.*, 2002). The microbial organisms transform the substance through metabolic or enzymatic processes (Bhandari, et al., 2021). It is based on two process-growth and co metabolism. In growth, an organic pollutant is used as sole source of carbon and energy. This process results in a complete degradation of organic pollutants. Cometabolism is defined as the metabolism of an organic compound in the presence of a growth substrate that is used as the primary carbon and energy source (Fritsche and Hofrichter, 2008). Several microorganisms like aerobic as well as anaerobic bacteria, algae and fungi are involved in biodegradation process (Das and Chandran, 2011). Biodegradation processes vary greatly, but frequently the final product of the degradation is carbon dioxide (Pramila, *et al.*, 2012).

www.garph.co.uk

ISSN: 2278-6252

Organic material can be degraded aerobically, with oxygen, or anaerobically, without oxygen (Mrozik, *et al.*, 2003; Fritsche and Hofrichter, 2008). The term biodegradation is often used in relation to ecology, waste management and mostly associated with environmental remediation. In the present review paper, the xenobiotic compounds are discussed and their biodegradation process in the environment are reviewed and summarized.

# **Type of Xenobiotic Compounds**

The recalcitrant xenobiotic compounds can be grouped into the following types:

#### **Halocarbons**

These compounds contain different numbers of halogen (Cl, Be, F and I) atoms in the place of H atoms. They are used as solvents (Chloroform, CHCl3), as propellants in spray cans of cosmetics, paints etc., in condenser units of cooling systems (Freons, CCl3F, CCl2F2, CClF3, CF4) and as insecticides (DDT, BHC, lindane) and herbicides (2, 4-D, 2, 4, 5-T and dalapon) Brillas, et al., 2003). The C1-C2 haloalkanes like chloroform, freons etc. are volatile and escape into the atmosphere where they destroy the protective ozone (O3) layer leading to increased UV radiation. Pesticides like herbicides, fungicides and insecticides are applied to crops from where they leach into water bodies; many of them are subject to biomagnifications.

## **Polychlorinated Biphenyls (PCB's)**

These compounds have two covalently linked benzene rings having halogens substituting for Polychlorinated biphenyls are used as plasticisers, insulator coolants in transformers and as heat exchange fluids (Taniguchi, *et al.*, 1997; Dhakal, et al., 2018). They are both biologically and chemically inert to various degrees, which increases with the number of chlorine atoms present in the molecules.

## **Synthetic Polymers**

These compounds are produced as plastics e.g. Polyethylene, polystyrene, polyvinyl chloride etc. and nylons, which are used as garments, wrapping materials (Boehm and Farrington, 1984) etc. They are recalcitrant mainly due to their insolubility in water and molecular size.

# **Alkylbenzyl Sulphonates**

These are surface-active detergents superior to soaps. The sulphonate group present at one end resists microbial degradation, while the other end (non-polar alkyl end) becomes recalcitrant if it is branched (Gordon, *et al.*, 2008).

ISSN: 2278-6252

#### **Oil Mixtures**

Oil is a natural product, has many components is biodegradable, the different components being degraded at different rates. Biodegradation is able to handle small oil seepages. But when large spills occur the problem of pollution becomes acute (Christensen, *et al.*, 2004; Hostettler, *et al.*, 2007). Oil is recalcitrant mainly because of its insolubility in water and due to toxicity of some of its components.

## **Other Xenobiotic Compounds**

A number of pesticides are based on aliphatic, cyclic ring structures containing substitution of nitro-sulphonate, methoxy, amino and carbamyl groups; in addition, they also contain halogens. These substitutions make them recalcitrant.

# **Hazard** of **Xenobiotic Compounds**

The xenobiotics present a number of potential hazards to human and the environment. These are following:

- Many
   xenobiotics like halogenated and aromatic hydrocarbons are toxic to bacteria, lower
   eukaryotes and even humans. At low concentrations they may cause various skin
   problems and reduce reproductive potential.
- 2. Certain halogenated hydrocarbons have been shown to be carcinogenic.
- 3. Many xenobiotics are recalcitrant and persist in the environment so that there is a build up in their concentration with timer.
- 4. Many xenobiotics including DDT and PCB's are recalcitrant and lipophilic; as a consequence they show bioaccumulation or biomagnifications often by a factor of  $10^{4-}$ - $10^{6}$  (Biomagnifications occurs mainly because of the following reasons:
  - (i) These compounds are continuously taken up from the environment and accumulation of DDT by phyto and zoo planktons from water.
  - organisms are consumed by other organisms in a sequential manner constituting the food chain e.g. Planktons→Small fishes→Large fishes→Seaeagles, the concentration of xenobiotics builds up as we move up in the food

ISSN: 2278-6252

chain (Iovdijova and Bencko, 2010; Godheja, *et al.*, 2016; Sall, *et al.*, 2022). In case of DDT a 10<sup>5</sup> fold increase occurs in Sea-eagles as compared to the concentration present in the aqueous environment as a result of which Sea-eagles laid fragile eggs. DDT and PCB's have been found in human tissues in high but sub lethal concentrations in those countries where they have been used, although humans were often not in direct contact with these chemicals (Cox and Surgan, 2006).

ISSN: 2278-6252

**Impact Factor:8.388** 

# **Degradation of Xenobiotic Compounds by Fungi**

Fungi are an important part of degrading microbiota because, like bacteria, they metabolize dissolved organic matter; they are principal organisms responsible for the decomposition of carbon in the biosphere (Harms, *et al.*, 2017). But, fungi, unlike bacteria, can grow in low moisture areas and in low pH solutions, which aids them in the breakdown of organic matter (Spellman, 2008). Equipped with extracellular multienzyme complexes, fungi are most efficient, especially in breaking down the natural polymeric compounds. By means of their hyphal systems they are also able to colonize and penetrate substrates rapidly and to transport and redistribute nutrients within their mycelium (Matavuly and Molitoris, 2009). Several yeasts may utilize aromatic compounds as growth substrates, but more important is their ability to convert aromatic substances co metabolically. Some species such as the soil yeast *Trichosporon cutaneum* possess specific energy-dependent uptake systems for aromatic substances (e.g. phenol) (Mörtberg and Neujahr, 1985; Benmessaoud, *et al.*, 2022).

Furthermore, biodegradation of aliphatic hydrocarbons occurring in crude oil and petroleum products has been investigated well, especially for yeasts. The n-alkanes are the most widely and readily utilized hydrocarbons, with those between C10 and C20 being most suitable as substrates for microfungi (Bartha, 1986). However, the biodegradation of n-alkanes having chain lengths upto n-C24 has also been demonstrated (Fritsche and Hofrichter, 2005). Typical representatives of alkane, utilizing yeasts include *Candida lipolytica, Candida tropicalis, Rhodotorula rubra* and *Aureobasidion (Trichosporon) pullulans. Rhodotorula aurantiaca* and *Candida ernobii* were found able to degrade diesel oil (De Ca`ssia Miranda, *et al.*, 2007). Yeasts are also reported for aniline biodegradation (Mucha, *et al.*, 2010). In addition to aromatic and aliphatic hydrocarbons compounds, fungi may transform numerous of other aromatic organopollutants co metabolically, including polycyclic aromatic hydrocarbons (PAHs) and biphenyls, dibenzofurans, nitroaromatics, various pesticides and plasticizers (Fritsche and Hofrichter, 2000; Atagana, 2009; Godoy, *et al.*, 2016; Zhang, *et al.*, 2017;

Wolf, et al., 2020; Correa, et al., 2021). There have also been studies of PCB metabolism by yeasts Candida boidinii and Candida lipolytica (Sasek, et al., 1993) and Saccharomyces cerevisiae (Eaton, 1985; Benmessaoud, et al., 2022). Insecticides and fungicides can also be adsorbed by Saccharomyces cerevisiae during aerobic fermentation (Cabras, et al., 1988; Spina, et al., 2018).

Yeasts are known for playing an important role in the removal of toxic heavy metals. There are many reports on biosorption of heavy metals by yeasts (Sharaf and Alharbi, 2013). Several investigations demonstrated that yeasts are capable of accumulating heavy metals such as Cu (II), Ni (II), Co (II) and Cd (II) and are superior metal accumulators compared to certain bacteria (Wang and Chen, 2006; Atagana, 2009; Gururajan and Belur, 2018). Several yeast strains like Saccharomyces cerevisiae, Rhodotorula pilimanae, Yarrowiali polytica, and Hansenula polymorpha have been reported to reduce Cr (III) (Ksheminska, et al., 2006). Most studies have reported the efficiency of immobilized cells of yeasts in metals removal. The majority of filamentous fungi are unable to totally mineralize aromatic hydrocarbons; they only transform them into indirect products of decreased toxicity and increased susceptibility to decomposition with the use of bacteria suggesting that the interaction among fungi and bacteria is profitable for the process of petroleum hydrocarbon mineralization. Among the filamentous fungi participating in aliphatic hydrocarbon are Cladosporium and Aspergillus, whereas fungi belonging to Cunninghamella, Penicillium, Fusarium and Aspergillus can take part in aromatic hydrocarbon decomposition (Steliga, 2012). Fungal genera, like Amorphoteca, Neosartorya and Talaromyces were isolated from petroleum contaminated soil and proved to be the potential organisms for hydrocarbon degradation (Chaillan and Bury, 2004). A group of fungi like Aspergillus, Cephalosporium and Penicillium was also found to be potential degrader of crude oil hydrocarbons (Singh, 2006). Fungal potentiality in PCBs degradation has been rarely explored. Several studies revealed that filamentous fungi can degrade PCBs. Among the filamentous fungi, the ligninolytic ones have been specifically investigated because of their extracellular, a specific oxido-reductive enzymes that have been already successfully exploited in the degradation of many aromatic pollutants. Fungi are known to tolerate and detoxify metals by several mechanisms including valence transformation, extra and intracellular precipitation and active uptake (Gadd, 1993; 2016). The most widely researched fungi in regard to dye degradation are the ligninolytic fungi (Bumpus, 2004). Nine strains of filamentous fungi were isolated by Abrusia, et, al., (2007) from cinematographic film consisted of three species of Aspergillus i.e. Aspergillus

ISSN: 2278-6252

ustus, Aspergillus nidulans, Aspergillus versicolor as well as Penicillium chrysogenum, Cladosporium cladosporioides, Alternaria alternate, Mucor racemosus, Phoma glomerata and Trichoderma longibrachiatum were able to biodegrade the gelatine emulsion with different rates of metabolic carbon dioxide production.

### **Conclusion**

In conclusion, xenobiotics include many compounds that are involved in both industrial and agricultural activities. Synthetic organic pesticides are a class of xenobiotics that are commonly used in agriculture and are added to the soil in large amount each year. Chemicals with pesticide activity were designed primarily to control insect, weed, fungi and nematode pests like DDT, BHC, 2, 4-D, 2, 4, 5-T, dalapon and carbomyl. The environmental conditions such as water content, salinity, pH and temperature must be suitable for the degrading microorganisms to proliferate and the enzyme to operate. If the xenobiotic compounds are toxic and the rate of biodegradation is very slow, adverse impact on humans, animals, fishes, birds and on ecological health are possible. It is describable to maintain xenobiotic concentrations in the environment at as low a level as possible. Because of these concerns, considerable research effort has been spent in trying to understand the metabolic degradation pathways of xenobiotic compounds in the environment. Several species of fungi play a major role for degradation of xenobiotic compounds in the environment. So, fungal activities are very important for the renewal of our environment and maintenance of the global carbon cycle.

#### References

Abruscia, C., Marquinna, D., Del Amob, A. and Catalina, F. (2007). Biodegradation of cinematographic gelatine emulsion by bacteria and filamentous fungi using indirect impedance technique. International Biodeterioration and Biodegradation, **60**: 137-

Akhtar, N. Iqbal, M., Iqbal, Z. S. and Iqbal, J. (2008). Biosorption characteristics of unicellular green alga *Chlorella sorokiniana* immobilized in loofa sponge for removal of Cr (III). *Journal of Environmental Sciences*, **20**: 231-239.

Alexander, M. (1965). Persistence and biological reactions of pesticides in soils. Soil Science Society of America, Proceedings, **29:** 1-7.

Alexander, M. (1980). Biodegradation of chemicals of environmental concern. *Science*, **211:** 132-138.

ISSN: 2278-6252

Atagana, H. I. (2009). Biodegradation of PAHs by fungi in contaminated-soil containing cadmium and nickel ions. *African Journal Biotechnology*, 8: 5780-5789.

Doi: 10.5897/AJB2009.000-9465

Bartha, R. (1986). Biotechnology of petroleum pollutant degradation. *Microbial Ecology*, **12:** 155-172.

Bennet, J.W., Wunch, K.G. and Faison, B.D. (2002). Use of fungi biodegradation. Manual of environmental microbiology, 2<sup>nd</sup> ed., ASM Press; Washington, D. C., 960-971.

Boehm, P.D. and Farrington, J.W. (1984). Aspects of the polycyclic aromatic hydrocarbon geochemistry of recent sediments in the Georges Bank region. *Environmental Science* and Technology, **18:** 840-845.

Bumpus, J. A. (2004). Biodegradation of azodyes by fungi. In: Arora, D. K. (Ed.) Fungal Biotechnology in Agriculture, Food and Environmental Applications. Marcel Dekker, New York, pp. 457-480.

Brillas, E., Calpe, J.C. and Cabot, P.L.(2003). Degradation of the herbicide 2, dichlorophenoxyacetic acid by ozonation catalyzed with Fe2p and UVA light. Applied Catalysis B: *Environmental*, **46:** 381-391.

Cabras, P., Meloni, M., Pirisi, F. M., Farris, G. A. and Faticheti, F. (1988). Yeast and pesticide interaction during aerobic fermentation. *Applied Microbiology* and *Biotechnology*, **29** (2-3): 298-301. Doi: 10.1007/BF01982920

Ceci, A., Pinzari, F., Russo, F., Persiani, A. M. and Gadd, G. M. (2019). Roles of saprotrophic fungi in biodegradation or transformation of organic and inorganic pollutants in co-contaminated sites. *Applied Microbiology and Biotechnology*, 103 (1): 53-68. https://doi.org/10.1007/s00253-018-9451-1

Cerniglia, C. E. and Gibson, D. T. (1977). Metabolism of naphthalene by *Cunnighamella elegans*. *Applied and Environmental Microbiology*; **34:** 363-370.

Chaillan, F., Le Flèche, A. and Bury, E. (2004). Identification and biodegradation potential of tropical aerobic hydrocarbon degrading microorganisms. *Research in Microbiology*, **155(7):** 587-595.

Chen, M., Shih, K., Hu, M., Li, F., Liu, C., Wu, W. and Tong, H. (2012). Biostimulation of Indigenous Microbial Communities for Anaerobic transformation of pentachlophenol in paddy soils of Southern China. *Journal of Agricultural and Food Chemistry*; **60:** 2967-2975.

www.garph.co.uk

ISSN: 2278-6252

Christensen, J.H., Hansen, A.B., Moretensen, J., Tomasi, G. and Andersen, A. (2004). Integrated methodology for forensic oil spill identification. *Environmental Science and Technology*, **38:** 2912-2918.

Cox, C. and Surgan, M. (2006). Unidentified inert ingredients in pesticides: Implications for human and environmental health. *Environmental Health Perspectives*, **114(12)**: 1803-

N. P. Das, and Chandran, (2011).Microbial degradation of petroleum hydrocarbons contaminants: An Overview SAGE-Hindawi Access to Research Biotechnology. Research International. Article ID 941810, 13Pages. 10.4061/2011/941810

De Cássia Miranda, R. and de Souza, C. S., de Barros Gomes, E., Lovaglio, R. B., Lopes, C. E. and de Fatima Vieira de Queiroz Sousa, M. (2007). Biodegradation of diesel oil by yeasts isolated from the Vicinity of Suape Port in the state of Pernambuco-Brazil.

\*\*Brazilian archives of biology and technology, 50 (1): 147-152.\*\*

Dwivedi, S. (2012): Bioremediation of heavy metal by algae: Current and future perspective. *Journal of Advanced Laboratory Research in Biology*, (3) 3.

Eaton, D. C. (1985). Mineralization of polychlorinated biphenyls by *Phanerochaete chrysosporium*: a ligninolytic fungus. Enzyme and Microbial Technology, **7:** 194-196.

El Fantroussi, S. and Agathos, S. N. (2005). Is bioaugmentation a feasible strategy for pollutant removal and site remediation? *Current Opinion in Microbiology*, **8:** 268-275.

Fritsche, W. and Hofrichter, M. (2008). Aerobic degradation by microorganisms. In Biotechnology Set, 2<sup>nd</sup> edition (Eds: H.J. Rehm and G. Reed), Wiley-VCH Verlag GmbH, Weinheim, Germany. Doi: 10.1002/9783527620999.ch6m

Fritsche, W. and Hofrichter, M. (2000). Aerobic degradation by microorganisms, In: Biotechnology. Vol 11b, Environmental Processes, (Eds: Rehm, H. J. and Reed, G.), Wiley-VCH, Weinheim, pp. 145-167.

Fritsche, W. and Hofrichter, M. (2005). Aerobic degradation of recalcitrant organic compounds by microorganisms, in environmental biotechnology: Concept and Applications (Eds: H. J. Jördening and J. Winter), Wiley-VCH Verlang GmbH & Co.KGaA, Weinheim, FRG. Doi: 10.1002/3527604286.ch7

Gadd, G. M. (1993). Interaction of fungi with toxic metals. New Phytologist, 124: 25-60.

ISSN: 2278-6252

Gadd, G. M. (2016). Fungi and Industrial Pollutants. In: C. P. Kubicek, I. S. Druzhinina (Eds.) The Mycota, volume IV: Environmental and Microbial relationships. Springer, Heidelberg, pp. 89-125.

Godoy, P., Reina, R., Calderón, A., Wittich, R-M., García-Romera, I. and Aranda, E. (2016). Exploring the potential of fungi isolated from PAH- polluted soil as a source of xenobiotics degrading fungi. *Environ. Sci. Pollut. Res.*, 23: 20985-20996. Doi: 10.1007/s//356-016-7257-1

Gordon, A.K., Blatch, G.L., Daniel, S. and Muller, W.J. (2008). Stresss protein responses in South African freshwater invertibrates exposed to detergent surfactant linear alkylbenzene sulfonate (LAS). Water Air and Soil Pollution, 193 (1-4): 123-130. Grishchenkov, V.G., Townsend, R.T., McDonald, T.J., Autenrieth, R.L., Bonner, J.S. and Boronin, A.M. (2000). Degradation of petroleum hydrocarbons by facultative anaerobic bacteria under aerobic and anaerobic conditions. Process Biochemistry; 35(9): 889-896.

Gururajan, K. and Belur, P. D. (2018). Screening and selection of indigenous metal tolerant fungal isolates for heavy metal removal. *Environ. Technol. Innov.*, **9:** 91-99. Doi: 10.1016/j.eti.2017.11.001

Harms, H., Wick, L. and Schlosser, D. (2017). The fungal community in organically polluted systems. In: Dighton, J. and White, J. F. (Eds.) The fungal community: its organization and role in the ecosystem, Fourth Edition. CRC Press/ Taylor and Francis Group, NY, USA, pp 459-469.

Hostettler, F.D., Wang, Y., Huang, Y.S., Cao, W.H., Bekins, B.A., Rostad, C.E., *et al.* (2007). Forensic fingerprinting of oil-spill hydrocarbons in a methanogenic environment-Mandan, ND and Bemidji, M.N. *Environmental Forensics*, **8(1-2):** 139-

Iovdijova, A. and Bencko, V. (2010). Potential risk of exposure to selected xenobiotic residues and their fate in the food chain-part I: classification of xenobiotics. *Annl. Agric. Environ. Med.*, 17: 183-192.

Jayaraj, R., Megha, P. and Sreedev, P. (2016). Organochlorine pesticides, their toxic effects on living organisms and their fate in the environment. *Interdiscip. Toxicol.*, **9:** 90-100. Doi: 10.1515/intox-2016-0012

ISSN: 2278-6252

Kafilzadeh, F., Sahragard, P., Jamali, H. and Tahery, Y. (2011). Isolation and identification of hydrocarbons degrading bacteria in soil around Shiraz Refinery.

\*\*African Journal of Microbiology Research, 4(19):3084-3089.

Kanade, S N., Adel, A. B. and Khilare, V. C. (2012). Melathion degradation by *Azospirillum lipoferum* Beijerinck. Science Research Reporter; **2** (1): 94-103.

Kobayashi, H. and Rittman, B. E. (1982). Microbial removal of hazardous organic compounds. *Environmental Science and Technology*; **16:** 170-183.

Ksheminska, H. P., Taras, M. H., Galyna, Z., Gayda, M. and Gonchar, V. (2006). Extracellular chromate reducing activity of the yeast cultures. *Central European Science Journals*, **1** (1): 137-149.

Ksheminska, H. P., Fedorovych, D., Honchar, T., Ivash, M. and Gonchar, M. (2008). Yeast tolerance chromium depends on extracellular chromate reduction and Cr (III) chelation. *Food Technology and Biotechnology*, **46 (4):** 419-426.

Marinescu, M., Dumitru, M. and Lacatusu, A. (2009). Biodegradation of Petroleum Hydrocarbons in an Artificial Polluted Soil. *Research Journal of Agricultural Science*, 41(2).

Matavuly, M. N. and Molitoris, H. P. (2009). Marine fungi degraders of poly-3-hydroxyalkanoate based plastic materials. Proceedings for Natural Sciences published by MaticaSrpska; **116:** 253-265.

Maurya, P. K. (2016). Bioaccumulation of xenobiotics compound of pesticides in riverine system and its control techniques: a critical review. *J. Ind. Poll. Control*, 32: 580-594.

Mishra, S., Lin,, Z., Pang, S., Zhang, W., Bhatt, P. and Chen, S. (2021). Recent Advanced Technologies for the characterization of Xenobiotic-Degrading Microorganisms and Microbial Communities. *Front. Bioeng. Biotechnol.*, **9:** 632059. Doi: 10.3389/fbioe.2021.632059

Mörtberg, M. and Neujahr, H. Y. (1985). Uptake of phenol in *Trichosporon* cutaneum. *Journal of Bacteriology*, **161:** 65-619.

Mrozik, A., Piotrowska-Seget, Z. and Labuzek, S. (2003). Bacterial degradation and bioremediation of Polycyclic Aromatic Hydrocarbons. *Polish Journal of Environmental Studies*, **12(1)**: 15-25.

www.garph.co.uk

ISSN: 2278-6252

Mrozik, A. and Piotrowska-Seget, Z. (2009). Bioaugmentation as a strategy for cleaning up of soils contaminated with aromatic compounds. *Microbial Research* 2006; Doi: 10.1016/j.micres.2009.08.001

Mucha, K., Kwapisz, E., Kucharska, U. and Okruszeki, A. (2010). Mechanism of aniline degradation by yeast strain *Candida methanosorbosa* BP-6. *Polish Journal of microbiology*, **59 (4):** 311-315.

Patowary, K., Patowary, R., Kalita, M. C. and Deka, S. (2016). Development of an efficient bacterial consortium for the potential remediation of hydrocarbons from contaminated sites. Front. Microbiol., 7: 1092. Doi: 10.3389/fmicb.2016.01092

Pena-Castro, J. M., Martinez-Jerónimo, F., Esparza-García, F. and Caňizares-Villanueva, R. O. (2004). Heavy metals removal by the microalga Scenedesmus incrassatulus in continuous cultures. *Bioresource Technology*, **94:** 219-222.

Petric, I., Hrak, D., Fingler, S., Vonina, E., Cetkovic, H., Begonja, A. K. and Udikovikoli, N. (2007). Enrichment and characterization of PCB-degrading bacteria as potential seed cultures for bioremediation of contaminated soil. *Food Technology and Biotechnology*: **45** (1): 11-20.

Pramila, R., Padmavathy, K., Ramesh, K. V. and Mahalakshmi (2012). Brevibacillus parabrevis, Acinetobacter baumannii and Pseudomonas citronellal is potential candidates for biodegradation of low density polyethylene (LDPE). Journal of Bacteriology Research, 4(1): 9-14.

Sasek, V., Volfova, O., Erbanova, P., Vyas, B. R. M. and Matucha, M. (1993). Degradation of PCBs by white rot fungi, methylotrophic and hydrocarbon utilizing yeasts and bacteria. *Biotechnology Letters*, **15**: 521-526.

Seeger, M., Camara, B. and Hofer, B. (2001). Dehalogenation, denitration, dehydroxylation and angular attack on substituted biphenyls and related compounds by dioxygenase. *Journal of Bacteriology*; **183**: 3548-3555.

Sharaf, E. F. and Alharbi, E. (2013). Removal of heavy metals from waste water of tanning leather industry by fungal species isolated from polluted soil. *Afr. J. Biotechnol.*, **12:** 4351-4355.

Singh, H. (2006). Mycoremedial: Fungal bioremediation, Wiley-Interscience, New York, NY, USA.

Spellman, F. R. (2008). Ecology for nonecologists. Page 176.

ISSN: 2278-6252

- Spina, F., Cecchi, G., Landinez-Torres, A., Pecoraro, L., Russo, F., Wu, B., Cai, L., Liu, X. Z., Tosi, S., Varese, G. C., Zotti, M. and Persiani, A. M. (2018). Fungi as a toolbox sustainable bioremediation of pesticides in soil and water. *Plant Biosyst*, **152**: 474-488. Doi: 10.1080/11263504.2018.1445130
- Steliga, T. (2012). Role of fungi in biodegradation of petroleum hydrocarbons in drill waste. *Polish Journal of Environmental studies*, **21 (2):** 471-479.
- Struthers, J. K., Jayachandran, K. and Moorman, T. B. (1998). Biodegradation of atrazine by *Agrobacterium radiobacter* J 14 a and use of this strain in bioremediation of contaminated soil. Applied of Environmental Microbiology; **64:** 3368-3375.
- Sunita, V. J., Dolly, P. R., Bateja, S. and Vivek, U. N. (2013). Isolation and screening for hydrocarbon utilizing bacteria (HUB) from petroleum samples. *Int, J. Curr. Appl. Sci.*, **2:** 48-60.
- Sun, G., Du, Y., Yin, J., Jiang, Y., Zhang, D., Jiang, B., et al., (2019). Response of microbial communities to different organochlorine pesticide (OCPs) contamination levels in contaminated soil. *Chemosphere*, **215**: 461-469. Doi: 10.1016/j.chemosphere.2018.09.160
- Surekha Rani, M., Lakshmi, V., Suvarnalatha Devi, K. P., Jaya Madhuri, R., Aruna, S., Jyothi, K., Narasimha, G. and Venkateswarlu, K. (2008). Isolation and characterization of a chlorpyrifos degrading bacterium from agricultural soil and its growth response. *African Journal of Microbiology Research;* (2): 26-31.
- Taniguchi, S., Murakami, A., Hosomi, M., Miymura, A. and Uchida, R. (1997). Chemical decontamination of PCB contaminated soil. *Chemosphere*, **34:** 1631-1637.
- Thierry, L., Armelle, B. and Karine, J. (2008). Performance of bioaugmentation-assisted phytoextraction applied to metal contaminated soils: A review. *Environmental Pollution*, **153**: 497-522.
- Walker, J. D., Colwell, R. R., Vaituzis, Z. and Meyer, S. A. (1975). Petroleum-degrading achlorophyllous alga *Protothecazopfi*. Nature (London); **254:** 423-424.
- Wang, J. and Chen, C. (2006). Biosorption of heavy metals by *Saccharomyces cerevisiae*: A Review. *Biotechnology Advances*, **24:** 427-451.
- Wang, X. C. and Zhao, H. M. (2007). Uptake and biodegradation of Polycyclic Aromatic Hydrocarbons by Marine Seaweed. (Proceedings of the 9<sup>th</sup> International Symposium). *Journal of Coastal Research*, SI **50**: 1056-1061.

ISSN: 2278-6252

Wiedemeier, T.M., Miller, R.N. and Wilson, J.T. (1995). Significance of anaerobic processes for the intrinsic bioremediation of fuel hydrocarbons: In, Proceedings of the Petroleum Hydrocarbons and Organic Chemicals in Groundwater-Prevention, Detection and Remediation Conference, Houston Texas.

Wolf, D. C., Cryder, Z., Khoury, R., Carlan, C. and Gan, J. (2020). Bioremediation of PAH-Contamination shooting range soil using integrated approaches. *Sci. Total Environ.*,
726: 13844. Doi: 10.1016/j.scitotenv.2020.138440

Zhang, S., Yao, H., Lu, Y., Yu, X., Wang, J., Sun, S., Liu, M., Li, Y-F and Zhang, D. (2017). Uptake and translocation of polycyclic aromatic hydrocarbons (PAHs) and heavy metals by maize from soil irrigated with wastewater. *Sci. Rep.* 7: 12165. Doi: 10.1038/s41598-01712437-w

Zhu, Y., Boye, A., Body-Malape, M. and Herkovits, J. (2017). The toxic effects of xenobiotics on the health of humans and animals. *Bio. Med. Res. Int.*, **17**: 4627872. Doi: 10.1155/2017/4627872

ISSN: 2278-6252